

Certificate of Analysis

pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

Part No.
E135A

Size
20µg

Part# 9PIE135

Revised 10/16



Instructions for use of this product can be found in the pGL4 Luciferase Reporter Vectors Technical Manual #TM259, available online at: www.promega.com/protocols/

Description: The pGL4.34[*luc2P*/SRF-RE/Hygro] Vector^(+/-) contains a Serum Response Factor Response Element (SRF-RE) that drives transcription of the luciferase reporter gene *luc2P* in response to activation of Serum Response Factor through multiple signaling pathways including activation of RhoA GTPase. *luc2P* is a synthetically derived luciferase sequence with humanized codon optimization. The *luc2P* gene also contains hPEST, a protein destabilization sequence. The protein encoded by *luc2P* responds more quickly to induction than the protein encoded by the *luc2* gene. The vector backbone contains an ampicillin resistance gene to allow selection in *E. coli* and the mammalian-selectable marker for hygromycin resistance.

Concentration: 1µg/µl.

GenBank® Accession Number: FJ773213.

Storage Buffer: The pGL4.34[*luc2P*/SRF-RE/Hygro] Vector is supplied in 10mM Tris-HCl, 1mM EDTA (pH 8.0).

Storage Conditions: See the Product Information Label for storage temperature recommendations and expiration date.

Usage Note: Mix well prior to use.



AF9PIE135 1016E135



Promega

Promega Corporation

2800 Woods Hollow Road	
Madison, WI 53711-5399	USA
Telephone	608-274-4330
Toll Free	800-356-9526
Fax	608-277-2516
Internet	www.promega.com

Quality Control Assays

Contaminant Assays

Contaminating Nucleic Acid Assay: RNA, single-stranded DNA and chromosomal DNA are not evident in a specified sample of this vector as determined by agarose gel electrophoresis.

Nuclease Assay: Following incubation of 1µg of this vector in Restriction Enzyme Buffer at 37°C for 16–24 hours, no evidence of nuclease activity is detected by agarose gel electrophoresis.

Physical Purity: $A_{260}/A_{280} \geq 1.80$, $A_{260}/A_{250} \geq 1.05$.

Functional Assays

Identity Assay: The vector has been sequenced completely and has 100% identity with the published sequence available at: www.promega.com/vectors

Restriction Digestion: The functional purity of this vector DNA is verified by complete digestion with several restriction enzymes at 37°C for 1 hour. Samples are examined by agarose gel electrophoresis, and cut and uncut vector DNA are compared with marker DNA.

PRODUCT USE LIMITATIONS, WARRANTY, DISCLAIMER

Promega manufactures products for a number of intended uses. Please refer to the product label for the intended use statements for specific products. Promega products contain chemicals which may be harmful if misused. Due care should be exercised with all Promega products to prevent direct human contact.

Each Promega product is shipped with documentation stating specifications and other technical information. Promega products are warranted to meet or exceed the stated specifications. Promega's sole obligation and the customer's sole remedy is limited to replacement of products free of charge in the event products fail to perform as warranted. Promega makes no other warranty of any kind whatsoever, and SPECIFICALLY DISCLAIMS AND EXCLUDES ALL OTHER WARRANTIES OF ANY KIND OR NATURE WHATSOEVER, DIRECTLY OR INDIRECTLY, EXPRESS OR IMPLIED, INCLUDING, WITHOUT LIMITATION, AS TO THE SUITABILITY, PRODUCTIVITY, DURABILITY, FITNESS FOR A PARTICULAR PURPOSE OR USE, MERCHANTABILITY, CONDITION, OR ANY OTHER MATTER WITH RESPECT TO PROMEGA PRODUCTS. In no event shall Promega be liable for claims for any other damages, whether direct, incidental, foreseeable, consequential, or special (including but not limited to loss of use, revenue or profit), whether based upon warranty, contract, tort (including negligence) or strict liability arising in connection with the sale or the failure of Promega products to perform in accordance with the stated specifications.

©BY USE OF THIS PRODUCT, RESEARCHER AGREES TO BE BOUND BY THE TERMS OF THIS LIMITED USE LABEL LICENSE.

Researchers shall have no right to modify or otherwise create variations of the nucleotide sequence of the luciferase gene except that researchers may (1) create fused gene sequences, and (2) insert and remove nucleic acid sequences in splicing research. No other use or transfer of this product or derivatives is authorized. Researchers must either (1) use luminescent assay reagents purchased from Promega for all determinations of luminescence activity of this product and its derivatives; or (2) contact Promega to obtain a license for use of the product. For any uses outside this label license, contact Promega for supply and licensing information. This product is for research use only; no commercial use is allowed. For a full copy of this label license, including the definition of "commercial use," go to: www.promega.com/LULL.

©U.S. Pat. No. 8,008,006 and European Pat. No. 1341808.

©U.S. Pat. No. 7,728,118.

© 2009, 2015, 2016 Promega Corporation. All Rights Reserved.

GenBank is a registered trademark of U.S. Department of Health and Human Services.

Dual-Glo and Dual-Luciferase are registered trademarks of Promega Corporation. Bright-Glo and ONE-Glo are trademarks of Promega Corporation.

Products may be covered by pending or issued patents or may have certain limitations. Please visit our Web site for more information.

All specifications are subject to change without prior notice.

Product claims are subject to change. Please contact Promega Technical Services or access the Promega online catalog for the most up-to-date information on Promega products.

Part# 9PIE135
Printed in USA Revised 10/16.

Signed by:

R. Wheeler, Quality Assurance



Features List and Map for the pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

SRF-RE	33–147
minimal promoter (minP)	180–210
<i>luc2P</i> reporter gene	243–2018
SV40 late poly(A) region	2058–2279
SV40 early enhancer/promoter	2327–2745
synthetic hygromycin (Hyg ^r) coding region	2770–3807
synthetic poly(A) signal	3831–3879
reporter vector primer 4 (RVprimer4) binding region	3946–3965
ColE1-derived plasmid replication origin	4203
synthetic β-lactamase (Amp ^r) coding region	4994–5854
synthetic poly(A) signal/transcriptional pause site	5959–6112
reporter vector primer 3 (RVprimer3) binding region	6061–6080

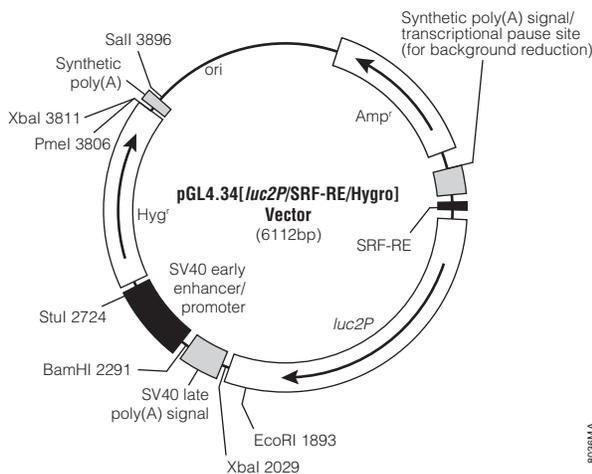


Figure 1. pGL4.34[*luc2P*/SRF-RE/Hygro] Vector map.

Sequence information, vector maps and restriction enzyme tables for the pGL4 Vectors are available online at: www.promega.com/vectors/

Additional information is available in Technical Manual #TM259, available online at: www.promega.com/protocols/

Sample Protocol to Determine Induction of Luciferase by FBS in HEK293 Cells Transfected with the pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

Materials to Be Supplied by the User

- Dulbecco's PBS (DPBS)
- 0.05% (w/v) trypsin in DPBS
- DMEM
- DMEM supplemented with , 0.5%, 10% and 40% fetal bovine serum (DMEM/FBS)
- ONE-Glo™ Luciferase Assay System (Cat.# E6110)
- HEK293 cells
- transfection reagent

Day 1: Plate Cells

1. Grow HEK293 cells in DMEM/FBS to approximately 75% confluency.
2. Harvest cells via trypsinization: Remove the DMEM/FBS, wash the cells with DPBS and add the trypsin/DPBS (1X volume). After 2 minutes, add a 4X volume of DMEM/FBS, collect the cell suspension and pellet the cells by centrifugation. Aspirate the supernatant, and resuspend in DMEM/FBS. We have routinely used a concentration of 10,000–15,000 viable cells/100µl DMEM/FBS.
3. Dispense 100µl of the cell suspension into the wells of a 96-well plate. Plate enough wells to perform each test condition in triplicate.
4. Cover the plate, and place it in a tissue culture incubator at 37°C overnight (or for 24 hours).

Day 2: Transfect Cells

1. Transfect the cells using a high-efficiency transfection reagent. Each well of cells in a 96-well plate requires 0.1µg pGL4.34[*luc2P*/SRF-RE/Hygro] Vector DNA. Transfection conditions may require optimization.
2. Cover the plate, and place it in a tissue culture incubator at 37°C.
3. After 4–6 hours, change the medium to DMEM/0.5%FBS (100µl per well) to start serum starvation.

Day 3: Induce Transfected Cells

1. Prepare 2X induction and 2X control solutions. Calculate the volume of 2X induction and 2X control solution by multiplying the number of wells needed for each solution by 50µl, and prepare 110% of this amount.
 - 2X induction solution: 40%FBS in DMEM
 - 2X control solution: DMEM
2. Remove 50µl of medium from wells that will be treated with either 2X induction solution or 2X control solution.
3. Add 50µl of 2X induction solution to the cells to be induced and 50µl of 2X control solution to the control noninduced cells.
4. Return the plate to the tissue culture incubator, and induce for 6 hours.
5. Analyze luciferase activity using an appropriate luciferase detection assay. We have observed comparable results for fold induction of the vector using a variety of luciferase reagents, including: Bright-Glo™ Luciferase Assay System (Cat.# E2610, Technical Manual #TM052); ONE-Glo™ Luciferase Assay System (Cat.# E6110, Technical Manual #TM292); Dual-Luciferase® Reporter Assay System (Cat.# E1910, Technical Manual #TM040); and Dual-Glo® Luciferase Assay System (Cat.# E2920, Technical Manual #TM058).
6. Calculate the fold induction as follows:

$$\text{fold induction} = \frac{\text{average relative light units of induced cells}}{\text{average relative light units of control cells}}$$