



**Promega**

# Technical Bulletin

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## **pAdVAntage™ Vector**

INSTRUCTIONS FOR USE OF PRODUCT E1711.



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Part# TB207

# pAdVAntage™ Vector

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## I. Description

Co-transfection of mammalian cells with the pAdVAntage™ Vector enhances transient protein expression in a variety of cell types by increasing translation initiation.

Transfection of mammalian cells with an expression vector often results in suboptimal expression of the protein of interest. Double-stranded RNA (dsRNA) generated during transfection is thought to activate the dsRNA-activated inhibitor (DAI), one of several enzymes involved in the host cell's antiviral defense system. DAI phosphorylates the translation initiation factor eIF-2, halting translation and therefore protein production (1, reviewed in reference 2; see Figure 1).

However, inhibition of translation by DAI can be overcome with the adenoviral Virus Associated I RNA (VAI RNA) that is produced by RNA polymerase III following co-transfection with the pAdVAntage™ Vector. The VAI RNA binds to DAI, preventing its activation, thereby allowing translation and protein expression (3,4, reviewed in reference 2; see Figure 1).

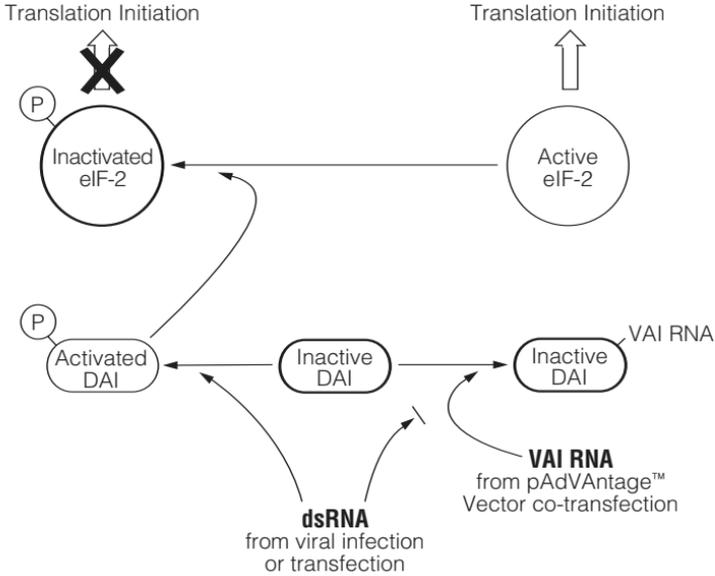
The pAdVAntage™ Vector contains base pairs 9,831–11,555 of the adenovirus type 2 genome on a 1,724bp SalI–HindIII fragment that encodes the virus-associated RNA genes, VAI and VAII.

## II. Product Components and Storage Conditions

Product	Size	Cat.#
pAdVantage™ Vector	20µg	E1711

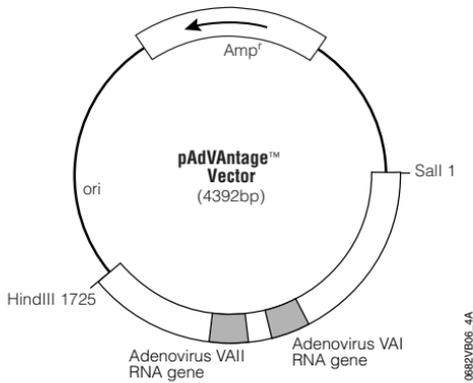
pAdVantage™ Vector is supplied frozen in TE buffer.

**Storage Conditions:** Store at -20°C.



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**Figure 1.** Translation initiation and the effects of VAI RNA on this process.



**Figure 2. pAdVantage™ Vector circle map and sequence reference points.**

**Sequence reference points:**

Adenovirus DNA insert	1–1725
Adenovirus VAI RNA gene	780–939
Adenovirus VAI RNA gene	1036–1198
Adenovirus VAI RNA gene	1036–1198
β-lactamase (Amp <sup>r</sup> ) coding region	2907–3764

### III. General Considerations

The level of expression enhancement attained by co-transfection with the pAdVantage™ Vector is dependent upon a number of parameters, including the cell type, method of transfection and the ratio of pAdVantage™ Vector DNA to expression vector DNA. Therefore, optimization of these parameters is critical for best results.

#### III.A. Optimization of the Ratio of pAdVantage™ Vector to Expression Vector DNA

To optimally enhance protein expression following co-transfection with the pAdVantage™ Vector, experimentally determine the optimal ratio of pAdVantage™ Vector to expression vector DNA for each transfection system. For initial optimization experiments, we recommend using ratios of pAdVantage™ Vector to expression vector DNA in the range of 1:10 to 1:1, as demonstrated in the sample in Table 1.

**Table 1. Sample Optimization of the Ratio of Co-Transfected DNAs.**

Transfected DNA	DNA Ratios (pAdVantage™ Vector DNA:Expression Vector DNA)			
	1:10	1:5	1:2.5	1:1
pAdVantage™ Vector DNA (or pBR322 DNA in control plates)	0.5µg	1µg	2µg	5µg
Expression Vector DNA	5µg	5µg	5µg	5µg

#### III.B. Sample Experiments Demonstrating pAdVantage™ Vector Enhancement of Expression in HeLa and 293 Cells

To determine how well the pAdVantage™ Vector can enhance protein expression, we performed co-transfection experiments of 293 and HeLa cells with pGL2-Control DNA and either pAdVantage™ DNA or pBR322 DNA as a control.

The cells were transfected in 60mm tissue culture dishes with 5µg of the pGL2-Control DNA (Cat.# E1611) using the ProFection® Mammalian Transfection System – Calcium Phosphate (Cat.# E1200). The ratio of pAdVantage™ DNA or pBR322 DNA co-transfected with the pGL2-Control DNA ranged from 1:10 to 1:1 as detailed in Table 1. The cells were lysed forty-eight hours post-transfection, and luciferase activity expressed from the pGL2-Control DNA was measured using the Luciferase Assay System (Cat.# E1500). The relative-fold increase in luciferase activity due to co-transfection with the pAdVantage™ Vector was determined using the luciferase activity of the control cells co-transfected with pBR322 as a baseline for comparison.

In these experiments, 293 cells co-transfected with the pAdVantage™ Vector demonstrated a 10- to 70-fold increase in luciferase expression from pGL2-Control DNA, while HeLa cells demonstrated a 4- to 40-fold increase in luciferase expression when co-transfected with the pAdVantage™ Vector.

#### IV. pAdVantage™ Vector Restriction Sites

The following restriction enzyme tables were constructed using DNASTAR® sequence analysis software. Please note that we have not verified this information by restriction digestion with each enzyme listed. The location given specifies the 3' end of the cut DNA (the base to the left of the cut site). For more information on the cut sites of these enzymes or if you identify a discrepancy, please contact your local Promega Branch or Distributor. In the U.S., contact Promega Technical Services at 800-356-9526. Vector sequences are also available from GenBank® database (GenBank®/EMBL Accession Number **U47294**) and on the Internet at: [www.promega.com/vectors/](http://www.promega.com/vectors/)

**Table 1. Restriction Enzymes That Cut the pAdVantage™ Vector Between 1 and 5 Times.**

Enzyme	# of Sites	Location	Enzyme	# of Sites	Location
<b>AatII</b>	2	914, 3899	<b>CspI</b>	1	1631
<b>AccI</b>	2	2, 314	<b>Csp45I</b>	1	841
<b>AccIII</b>	1	1171	<b>DraI</b>	3	2843, 2862, 3554
<b>Acc65I</b>	1	4372	<b>DraII</b>	3	1084, 1407, 3953
<b>AflIII</b>	2	1656, 2084	<b>DrdI</b>	2	2192, 4061
<b>Alw44I</b>	3	2398, 3644, 4141	<b>DsaI</b>	5	701, 790, 1325, 1363, 1489
<b>AlwNI</b>	3	494, 1500, 2500	<b>EagI</b>	3	855, 1108, 1664
<b>Apal</b>	1	1377	<b>EarI</b>	5	215, 791, 1968, 3772, 4260
<b>AvaI</b>	3	1101, 1528, 4376	<b>EclHKI</b>	2	321, 2977
<b>BalI</b>	2	382, 982	<b>Eco47III</b>	1	930
<b>BamHI</b>	2	850, 4381	<b>Eco52I</b>	3	855, 1108, 1664
<b>BbeI</b>	3	207, 1371, 4203	<b>Eco72I</b>	1	1659
<b>BbrPI</b>	1	1659	<b>EcoICRI</b>	2	425, 4368
<b>BbsI</b>	1	566	<b>EcoNI</b>	1	306
<b>BglII</b>	4	265, 726, 3097, 4215	<b>EcoRI</b>	1	4360
<b>BsaI</b>	1	3038	<b>EcoRV</b>	2	499, 595
<b>BsaAI</b>	2	1485, 1659	<b>EheI</b>	3	205, 1369, 4201
<b>BsaMI</b>	1	44	<b>FseI</b>	1	1110
<b>BsmI</b>	1	44	<b>FspI</b>	5	43, 682, 734, 3199 4222
<b>Bsp120I</b>	1	1373	<b>HincII</b>	3	3, 398, 743
<b>BspHI</b>	3	2804, 3812, 3917	<b>HindII</b>	3	3, 398, 743
<b>BspMI</b>	2	77, 310	<b>HindIII</b>	1	1725
<b>BssHII</b>	4	646, 992, 1648, 1650	<b>HpaI</b>	1	398
<b>BssSI</b>	3	2257, 3641, 3948	<b>KasI</b>	3	203, 1367, 4199
<b>BstEII</b>	3	407, 878, 1677	<b>KpnI</b>	1	4376
<b>BstXI</b>	1	21	<b>NaeI</b>	2	629, 1108
<b>BstZI</b>	3	855, 1108, 1664	<b>NarI</b>	3	204, 1368, 4200
<b>Cfr10I</b>	5	121, 627, 720, 1106, 3057	<b>NcoI</b>	1	701

**Note:** The enzymes listed in boldface type are available from Promega.

#### IV. pAdVantage™ Vector Restriction Sites (continued)

**Table 1. Restriction Enzymes That Cut the pAdVantage™ Vector Between 1 and 5 Times (continued).**

Enzyme	# of Sites	Location	Enzyme	# of Sites	Location
NdeI	1	4148	SacI	2	427, 4370
NgoMIV	2	627, 1106	SacII	3	1328, 1366, 1492
NheI	1	969	Sall	1	1
NotI	1	1664	ScaI	1	3457
NruI	1	1508	SmaI	1	4378
NsiI	1	1214	SplI	2	1481, 1685
NspI	3	1280, 2088, 4005	SspI	1	3781
PmlI	1	1659	StyI	4	20, 701, 1067, 1447
Ppu10I	1	1210	TfiI	2	1919, 2059
PpuMI	1	1084	Tth111I	1	468
Psp5II	1	1084	VspI	3	1855, 1914, 3149
PspAI	1	4376	XbaI	2	749, 4387
PvuI	2	3347, 4243	XmaI	1	4376
PvuII	3	1458, 1908, 4272	XmnI	1	3576
RsrII	1	1631			

**Table 2. Restriction Enzymes That Do Not Cut the pAdVantage™ Vector.**

AccB7I	BlpI	DraIII	PmeI	SphI
AflIII	Bpu1102I	Eco81I	PshAI	SrfI
AgeI	BsaBI	<b>I-PpoI</b>	<b>PstI</b>	Sse8387I
AscI	BsrGI	<b>MluI</b>	<b>SfiI</b>	<b>StuI</b>
AvrII	Bst1107I	PacI	<b>SgfI</b>	Swal
<b>BbuI</b>	<b>Bst98I</b>	PaeR7I	SgrAI	XcmI
<b>BclI</b>	<b>Bsu36I</b>	PfIMI	<b>SnaBI</b>	<b>XhoI</b>
<b>BglII</b>	<b>ClaI</b>	PinAI	<b>SpeI</b>	

**Table 3. Restriction Enzymes That Cut the pAdVantage™ Vector 6 or More Times.**

Acil	<b>Bsp1286I</b>	<b>FokI</b>	MaeIII	<b>Sau3AI</b>
AcyI	BsrI	<b>HaeII</b>	<b>MboI</b>	Sau96I
<b>AluI</b>	<b>BsrSI</b>	<b>HaeIII</b>	MnlI	ScrFI
<b>Alw26I</b>	Bst71I	HgaI	MseI	SfaNI
AspHI	<b>BstOI</b>	<b>HhaI</b>	<b>MspI</b>	<b>SinI</b>
<b>AvaII</b>	BstUI	<b>HinFI</b>	<b>MspAII</b>	<b>TaqI</b>
<b>BanI</b>	<b>CfoI</b>	<b>HpaII</b>	<b>NciI</b>	<b>Tru9I</b>
<b>BanII</b>	<b>DdeI</b>	HphI	<b>NdeII</b>	<b>XhoII</b>
BbvI	<b>DpnI</b>	<b>Hsp92I</b>	NlaIII	
BsaOI	DpnII	<b>Hsp92II</b>	NlaIV	
BsaHI	EaeI	MaeI	PleI	
BsaJI	Fnu4HI	MaeII	<b>RsaI</b>	

**Note:** The enzymes listed in boldface type are available from Promega.

## V. Related Products

Product	Size	Cat.#
ProFection® Mammalian Transfection System – Calcium Phosphate	40 transfections	E1200
ProFection® Mammalian Transfection System – DEAE-Dextran	40 transfections	E1210
Transfectam® Reagent for the Transfection of Eukaryotic Cells	1mg	E1231
	0.5mg	E1232
TransFast™ Transfection Reagent	1.2mg	E2431
Tfx™-20 Reagent	4.8mg	E2391
Tfx™-50 Reagent	2.1mg	E1811
Tfx™ Reagents Transfection Trio	5.4mg	E2400
pSI Mammalian Expression Vector	20µg	E1721
pCI Mammalian Expression Vector	20µg	E1731
pCI-neo Mammalian Expression Vector	20µg	E1841
pTARGET™ Mammalian Expression Vector	20 reactions	A1410

## VI. References

1. Farrell, P.J. *et al.* (1977) Phosphorylation of initiation factor eIF-2 and the control of reticulocyte protein synthesis. *Cell* **11**, 187-200.
2. Groskreutz, D. and Schenborn, E. (1994) Increased gene expression in mammalian cell lines using pAdVAntage™ DNA as a co-transfectant. *Promega Notes* **48**, 8-12.
3. Kitajewski, J. *et al.* (1986) Adenovirus VAI RNA antagonizes the antiviral action of interferon by preventing activation of the interferon-induced eIF-2 alpha kinase. *Cell* **45**, 195-200.
4. O'Malley, R. *et al.* (1986) A mechanism for the control of protein synthesis by adenovirus VA RNAI. *Cell* **44**, 391-400.

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